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Comparative Evaluation of Inductively Coupled Plasma-Optical Emission Spectrometry and Colorimetry for Determining Phosphorus in Grain Samples

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ABSTRACT

The inductively coupled plasma (ICP)-based method provides the opportunity to determine phosphorus (P) along with other major, secondary, micro and trace elements in plant materials. This study was conducted to compare and evaluate the relative efficacy of the inductively coupled plasma-optical emission spectrometry (ICP-OES) method with that of the colorimetric method using Skalar autoanalyzer, for determining P in 428 grain samples of eight diverse crops. The results on grain P analysis by the two methods, for individual as well as for all crop samples combined, showed that they were highly positively correlated (r varied from 0.84 to 0.98, $p < 0.0001$ for the eight crops, and R^2 for all crop grain samples was 0.9201, $p < 0.0001$). Moreover, the precision by the ICP method was similar to that determined by the Skalar method. Our results demonstrate that the ICP-OES method can be conveniently used for determining P along with other plant nutrient elements in grain samples of diverse crops.

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KEYWORDS

Colorimetric method;
ICP-OES; phosphorus in
grain; precision

Introduction

Phosphorus (P) deficiency is especially widespread in the soils of tropical regions of the world (Sahrawat et al. 2001; Bationo et al. 2008). In the ICRISAT analytical service laboratory, P in plant materials, including grain samples of various crops, is routinely determined by the colorimetric method using a Skalar autoanalyzer. However, with the availability of the inductively coupled plasma-optical emission spectrometer (ICP-OES) in our laboratory, we have an opportunity to determine P in plant samples by using ICP-OES as well. Indeed, this provides a very convenient method to determine P along with all other major, micro and trace elements in plant materials including grain samples. In addition, we wanted to evaluate the results obtained by the ICP-OES-based method with the standard colorimetric method using a Skalar autoanalyzer.

In earlier studies, we have observed that in general the ICP-AES (atomic emission spectrometer) method was far more sensitive than the turbidimetric method for the determination of extractable sulfur (S) (Shirisha et al. 2010), and the ICP-OES method was more sensitive than the colorimetric method for determining extractable boron (B) (Sahrawat et al. 2012) in diverse soils. The objective of this study, therefore, was to comparatively evaluate the Skalar and ICP-OES-based methods for determining the total P in grain samples of diverse crops including cereals, legumes and oilseeds.

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Materials and methods

Grain Samples

A total of 428 grain samples were selected to include a range of crops varying from cereals (pearl millet, sorghum, maize and rice), pulses (pigeonpea and chickpea) and oilseed (groundnut and sunflower) crops. The grain samples were dried at 60°C for 48 h before grinding the samples for total P analysis by the ICP-OES and Skalar methods.

Determination of P in grain sample by ICP-OES

Finely ground, dried grain samples were weighed (0.30 g) in 50 ml polypropylene (PP) tubes. A total of 2 ml of concentrated nitric acid (HNO₃) and 0.5 ml of 30% hydrogen peroxide (H₂O₂) (w/v) solution were added in that sequence, in each tube. Along with the samples, a number of blanks were included (with HNO₃ acid and H₂O₂). After properly mixing the contents in the tubes, they were kept for cold digestion overnight at room temperature (25°C). Next morning, the contents in the tubes were mixed, and placed in DigiPrep digestion blocks (SCP Scientific, Quebec, Canada) with a set temperature programming (room temperature to 80°C in 30 min, hold time 1 h, and then in 15 min rise of temperature from 80°C to 125°C and final hold time 2 h). The tubes were cooled and volume made up to 25 ml with distilled water. The contents of the tubes were mixed thoroughly for 1 min. The solution was filtered through Whatman filter paper number 1, and an aliquot from the digest was used for phosphorous analysis in ICP-OES (Prodigy, Teledyne Leeman, Hudson, N. H., USA). Detailed description of the method used is provided in Wheal, Fowles, and Palmer (2011).

Determination of P in Grain Sample by Skalar

Finely ground grain samples (0.3 g) were digested in 75 ml digestion tubes by adding 2.5 ml of concentrated sulfuric acid-selenium (Se) mixture (sulfuric acid containing 0.4% Se, v/w, was heated to dissolve Se) (Sahrawat, Ravi Kumar, and Murthy 2002), and P in the digests was analyzed using a Skalar autoanalyzer. All the grain samples were analyzed in three replications, and the results presented are the means of three independent analyses.

The data were statistically analyzed, and significance of the results by the two methods for P analysis was also tested. Correlations between the values of total P in grain samples by the ICP and Skalar methods were also determined.

Results and discussion

The results of the P concentration in grain samples of eight crops determined by the ICP-OES method were highly positively correlated with the values determined by the Skalar colorimetric method; moreover, the value of correlation coefficient *r* varied from 0.84 to 0.98 ($p < 0.0001$) for the eight crops (Table 1).

Table 1. Relationship between P (%) measured by Skalar autoanalyzer (Skalar-P) and inductively coupled plasma- optical emission spectrometer (ICP-P) in grain samples of eight crops.

Crops	Number of samples (n)	Regression equation	Correlation coefficient (<i>r</i>)	<i>p</i> -value
Pearl millet	28	Skalar-P = 0.0259 + 0.8878 ICP-P	0.88**	<0.0001
Maize	44	Skalar-P = 0.0505 + 0.8083 ICP-P	0.88**	<0.0001
Rice	45	Skalar-P = 0.0021 + 0.9847 ICP-P	0.98**	<0.0001
Sorghum	74	Skalar-P = 0.0462 + 0.7447 ICP-P	0.84**	<0.0001
Chickpea	50	Skalar-P = 0.0095 + 0.9404 ICP-P	0.95**	<0.0001
Pigeonpea	58	Skalar-P = -0.0002 + 0.9591 ICP-P	0.95**	<0.0001
Groundnut	91	Skalar-P = 0.0725 + 0.7683 ICP-P	0.85**	<0.0001
Sunflower	38	Skalar-P = 0.0569 + 0.8276 ICP-P	0.93**	<0.0001

**1% level of significance.

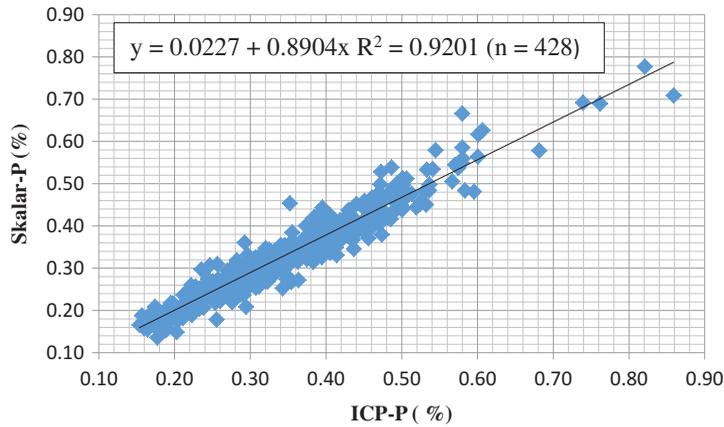


Figure 1. Relationship between Skalar-P and ICP-P in 428 grain samples of eight crops.

The relationship between the values of P concentrations determined by the ICP-OES method was highly positively correlated with that determined by the Skalar colorimetric method, considering all grain samples of all eight crops (Figure 1); the results are represented by the following regression equation:

$$\text{Skalar-P}(\%) = 0.0227 + 0.890 \text{ ICP-P}(\%), R^2 = 0.9201 (n = 428) \quad (1)$$

The precision obtained in determining total P in grain samples by the ICP and Skalar methods, as judged by the range, mean and standard deviation (SD) values, was similar and comparable for grain samples of eight crops individually as well as for all crops combined (Table 2).

Table 2. Precision obtained in the determination of P in grain samples of seven crops by the ICP (ICP-P) and Skalar (Skalar-P) methods for determining P in grain samples of eight crops.

Crops		ICP-P (%)	Skalar-P (%)
Pearl millet	Range	0.16–0.29	0.16–0.31
	Mean	0.22	0.22
	SD	0.04	0.04
Maize	Range	0.17–0.38	0.16–0.35
	Mean	0.26	0.26
	SD	0.05	0.04
Rice	Range	0.18–0.38	0.16–0.37
	Mean	0.30	0.30
	SD	0.04	0.04
Sorghum	Range	0.15–0.47	0.14–0.38
	Mean	0.27	0.25
	SD	0.06	0.05
Chickpea	Range	0.20–0.74	0.21–0.69
	Mean	0.40	0.40
	SD	0.11	0.11
Pigeonpea	Range	0.26–0.58	0.23–0.56
	Mean	0.38	0.36
	SD	0.07	0.07
Groundnut	Range	0.28–0.57	0.26–0.51
	Mean	0.42	0.40
	SD	0.06	0.05
Sunflower	Range	0.32–0.86	0.27–0.78
	Mean	0.49	0.46
	SD	0.13	0.11
All crops	Range	0.15–0.86	0.14–0.78
	Mean	0.30	0.29
	SD	0.11	0.10

Our results on the analysis of total P in grain samples of eight diverse crops by the ICP-OES method suggest that the method can be conveniently used for the determination of total P in grain samples of crops along with other plant nutrient elements. Indeed, the ICP-based methods provide an opportunity to determine P along with other major, micro and trace nutrient elements in diverse plant materials.

References

- Bationo, A., J. Kihara, B. Vanlauwe, J. Kimetu, B. S. Vaswa, and K. L. Sahrawat. 2008. Integrated nutrient management concepts and experience from Sub-Saharan Africa. In *Integrated nutrient management for sustainable crop production*, eds. M. S. Aulakh, and C. A. Grant, 467–521. New York, USA: The Haworth Press, Taylor and Francis Group.
- Sahrawat, K. L., M. K. Abekoe, and S. Diatta. 2001. Application of inorganic phosphorus fertilizer. In *Sustaining fertility in West Africa. Soil Science Society of America Special Publication no. 58*, eds. G. Tian, F. Ishida, and D. Keatinge, 225–46. Madison, WI, USA: Soil Science Society of America and American Society of Agronomy.
- Sahrawat, K. L., G. Ravi Kumar, and K. V. S. Murthy. 2002. Sulfuric acid-selenium digestion for multi-element analysis in a single plant digest. *Communications in Soil Science and Plant Analysis* 33:3757–65. doi:10.1081/CSS-120015920.
- Sahrawat, K. L., K. Shirisha, K. V. S. Murthy, P. N. Gajbhiye, S. Kundu, S. P. Wani, and G. Pardhasaradhi. 2012. Comparative evaluation of inductively coupled plasma-atomic emission spectroscopy and colorimetric methods for determining hot water-extractable boron in soils. *Communications in Soil Science and Plant Analysis* 43:948–57. doi:10.1080/00103624.2012.653030.
- Shirisha, K., K. L. Sahrawat, K. V. S. Murthy, S. P. Wani, P. N. Gajbhiye, and S. Kundu. 2010. Comparative evaluation of ICP-AES and turbidimetric methods for determining extractable sulfur in soils. *Journal Indian Society of Soil Science* 58:323–26.
- Wheal, M. S., T. O. Fowles, and L. T. Palmer. 2011. A cost-effective acid digestion method using closed polypropylene tubes for inductively coupled plasma optical emission spectrometry (ICP-OES) analysis of plant essential elements. *Analytical Methods* 3:2854–63. doi:10.1039/c1ay05430a.